PRACTICAL DERMATOLOGY

[Translated article] Dermoscopy of Squamous Cell Carcinoma: From Actinic Keratosis to Invasive Forms

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Abstract When the dermoscopy of squamous cell carcinoma and its precursors we differentiate among keratin-related, vascular, and pigment-related criteria. Non-pigmented actinic keratoses are characterized by the ''strawberry pattern''. Pigmented actinic keratosis shows a significant dermatoscopic overlap with lentigo maligna, but the presence of pigmented scales, erythema, and prominent follicles favors its diagnosis. Bowen's disease is characterized by clustered glomerular vessels, white-yellowish scales, and brown or grey dots arranged in lines in its pigmented variant. Finally, dermoscopy allows us to detect invasive squamous cell carcinoma in its early stages and differentiate it from its precursors. Furthermore, its presentation may vary depending on the degree of differentiation, with keratin-associated criteria predominating in well-differentiated tumors, while an atypical vascular pattern will predominate in poorly differentiated tumors.

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Dermatoscopia del carcinoma epidermoide: de la queratosis actínica a las formas invasivas

Resumen En la evaluación dermatoscópica del carcinoma epidermoide y sus precursores diferenciaremos entre criterios relacionados con la queratina, criterios vasculares y criterios relacionados con el pigmento. Las queratosis actínicas no pigmentadas se caracterizan por el denominado ''patrón en fresa''. Las queratosis actínicas pigmentadas presentan un gran solapamiento con el léntigo maligno, pero la presencia de escamas pigmentadas, el eritema y los folículos prominentes favorecen su diagnóstico. La enfermedad de Bowen se caracteriza por la presencia de agregados de vasos glomerulares y escamas blanco-amarillentas, así como por

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puntos marrones o grises dispuestos en líneas en su variante pigmentada. Por último, la dermatoscopia puede permitirnos la detección del carcinoma epidermoide invasivo en sus fases incipientes y diferenciarlo de sus precursores. Además, este variará en su presentación en función del grado de diferenciación, predominando los criterios asociados a la queratina en tumores bien diferenciados, mientras que en tumores mal diferenciados predominará un patrón vascular atípico.

Introduction

Squamous cell carcinoma (SCC) is the second most frequent skin cancer, representing approximately 20% of all non-melanoma skin cancers (NMSC). The incidence of SCC is on the rise.[1,2] The overall incidence rate in our region has been estimated at 38.16/100,000 persons/year.[3] Although most SCCs evolve satisfactorily after surgical excision, there is a subgroup of high-risk lesions with a high probability of recurrence, metastasis, and disease-related death.[1,3] Considering the progressive aging of the population and the corresponding increase in the incidence of NMSC—especially keratinocyte carcinomas—strategies aimed at the early diagnosis of malignant lesions and their differentiation from precursors are becoming increasingly important.

Dermoscopy is a non-invasive technique that is now an essential part of clinical diagnosis in dermatology. In experienced hands, it has been shown to improve diagnostic accuracy in both pigmented and non-pigmented skin lesions. Specifically, dermoscopy increases sensitivity in the diagnosis of SCC,[4] with various patterns associated with different types of lesions and stages of progression having been described, which facilitates differentiating among actinic keratosis (AK), Bowen's disease (BD), and invasive SCC (iSCC)[5,6] (Fig. 1). Similarly, pigmented AK (pAK) can pose diagnostic challenges due to its clinical overlap with lentigo maligna (LM), a scenario in which dermoscopy has also demonstrated its validity.[5] This review aims to synthesize the existing literature with a practical approach, addressing the spectrum of keratinocytic neoplasms and their precursors from a dermoscopic perspective while unifying the considerable terminological heterogeneity existing in Spanish[7] (Table 1).

This review will exclude SCC of the nail apparatus, which, due to its peculiarities and specific dermoscopic criteria, should be addressed in a separate work.

Dermoscopy of actinic keratosis

Non-pigmented actinic keratosis

Key aspects

Non-pigmented AK is frequently characterized by the so-called ''strawberry pattern,'' featuring a red pseudonetwork interrupted by prominent follicles. These can appear as ''rosettes'' under polarized light dermoscopy (Fig. 2).

In an initial trial, Zalaudek et al. studied a total of 41 non-pigmented facial AKs and identified a total of 4 fundamental dermocopic structures: a red pseudonetwork (95%), superficial scales (85%), fine linear or wavy perifollicular vessels (81%), and prominent follicles (66%), and/or surrounded by a white halo (''targetoid'') (100%). The combination of these structures forms the metaphorically termed ''strawberry pattern''.[8,9] This pattern was later observed by the same group in 67% of lesions in their series, and is significantly associated with the diagnosis of AK vs BD/SCC/keratoacanthoma (KA) (p < 0.001).[10]

Cuellar et al. described structures called ''rosettes'' (4 bright white dots resembling a ''four-leaf clover'') in AK, only visible with polarized light[11] (Fig. 2). However, these structures have subsequently been described in a wide range of neoplasms and even in non-lesional photo-damaged skin, which is why they are not considered specific.[12] Lozano-Masdemont et al. later proposed that the ''rosette pattern,'' present in 35.8% of lesions in their series characterized by this structure as the predominant feature could, indeed, could be specific.[13]

From a practical standpoint and according to Olsen's clinical classification, we will be observing these structures with relative frequency, which will eventually allow us to classify AKs into 3 clinical/dermoscopic stages[5] (Fig. 2):

Grade 1 AK: palpable, scarcely visible lesions characterized by a red pseudonetwork and discrete scaling under dermoscopy.

Grade 2 AK: visible, easily palpable, moderately keratotic lesions characterized by a reddish background with prominent follicles or ''rosettes'' under dermoscopy. This stage corresponds to the previously described ''strawberry pattern''[5].

Grade 3 AK: thick lesions with marked hyperkeratosis and well-demarcated borders, predominantly showing compact keratin masses as white/yellowish structureless areas under dermoscopy[5,14].

Regarding locations, we should mention that non-pigmented extrafacial AKs may present certain dermoscopic differences. In their study, Reinehr et al. saw that whitish scales (97.3%) and erythema (57.4%) were the most common structures. The anatomical differences described in extrafacial skin (primarily the lower density of adnexal structures) imply that follicle-associated structures, and the red pseudonetwork are less common findings.[15]

The validity of dermoscopy for diagnosing AKs was confirmed by Huerta-Brogueras et al. in a prospective trial of 178 clinically suggestive AK lesions, with sensitivity
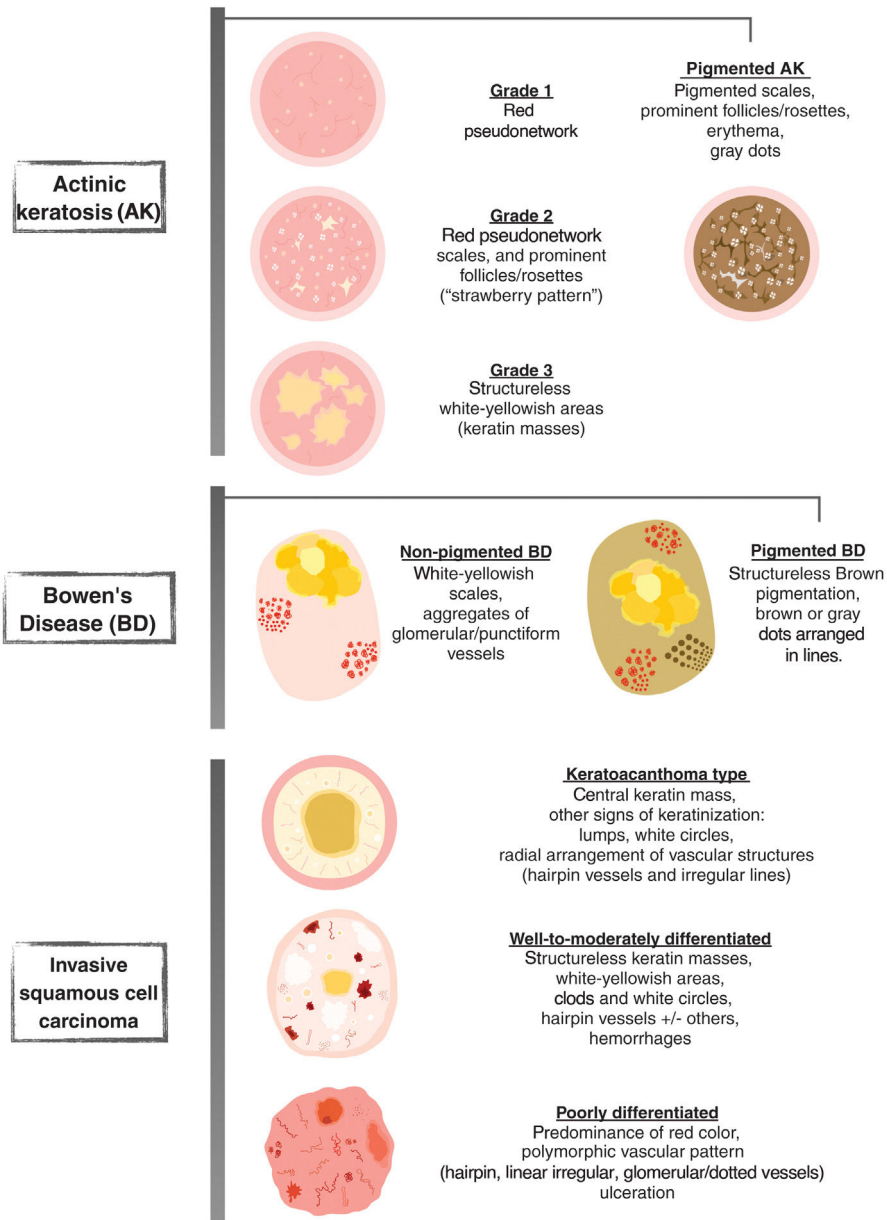
Figure 1 Schematic representation of dermoscopy of actinic keratoses, Bowen's disease, and keratoacanthoma/invasive squamous cell carcinoma.

and specificity rates of 98.7% and 95%, respectively, and a concordance of κ = 0.917 between this technique and histopathology.[16] Furthermore, dermoscopy also seems to be useful for post-therapeutic follow-up with cryotherapy, topical therapies, or photodynamic therapy (PDT).[17,18]

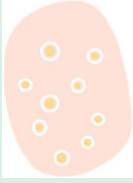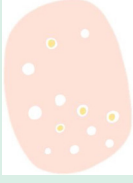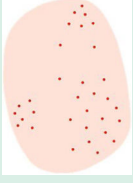## Pigmented actinic keratosis

Key aspects
pAK and LM present significant dermoscopic overlapping, primarily based on pigment-related criteria such as gray dots, rhomboidal structures, or asymmetric perifollicular pigmentation.

Additional findings, such as prominent follicles/''rosettes,'' the presence of scales, erythema, or an inner gray halo at follicular level favor the diagnosis of pAK (Fig. 3).

Diagnosing pAK can be challenging due to its clinical and dermoscopic similarity with LM[19–22]. In this regard, Akay et al. studied a total of 99 pigmented facial lesions (67 of them pAKs) and observed that the latter could exhibit dermoscopic criteria, such as gray dots (70%), an annular/granular pattern (39%), rhomboidal structures (36%), or asymmetric perifollicular pigmentation (25%).[23] The study by Moscarella et al. with 17 facial and extrafacial pAKs revealed that the most common structures were gray dots (76.5%), structureless brown areas (58.8%), pigmented pseudonetwork (35.3%), and the presence of white circles in 11.7% of the cases. Same as LM, these authors observed an annular/granular pattern and asymmetric perifollicular pig-

Table 1 Glossary of the main dermoscopic terms described in the literature in the spectrum of actinic keratosis, Bowen's disease, and invasive squamous cell carcinoma, along with their definition, schematic representation, and histopathological correlation.

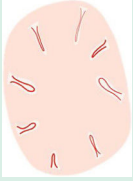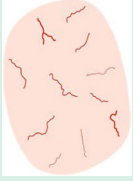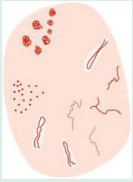| Criterion	Definition	Histopathological correlation	Schematic representation
Keratin-related criteria			
Superficial white-yellow or brown scales	Structureless white-yellow or brown opaque areas	Areas of hyperkeratosis and parakeratosis	
White clods (''keratin pearls'')	Rounded white-yellowish structures surrounded by a whitish halo	Keratin pearls or horn swirls	
White circles	Concentric white structures surrounding a follicular orifice, which may have a central yellow globular area	Acanthosis and hypergranulosis of the infundibular epidermis with a central keratin plug	
White structureless areas	Homogeneous areas that may cover a large part of the tumor, possibly associated with other white structures (circles, clods)	Hyperkeratosis and parakeratosis over a dysplastic epidermis or keratin in neoplastic cell aggregates	
''Rosettes''	4 bright white dots arranged like a ''4-leaf clover''	Optical effect of cross-polarization, resulting from alternating hyperkeratosis and parakeratosis in follicular orifices and/or concentric fibrosis	
Vascular criteria			
Red pseudonetwork	Structureless erythematous areas and wavy vessels surrounding follicular orifices	Localized increase in vascularization along with variable follicular hyperkeratosis and keratinocyte atypia	See Fig. 1
''Strawberry pattern''	Reddish pseudo-reticulum interrupted by prominent follicular orifices		See Fig. 1
Dotted vessels	Small caliber red dots resembling pinheads, densely packed together	Tips of short capillary loops in the papillary dermis	

Table 1 (*Continued*)
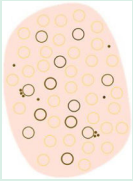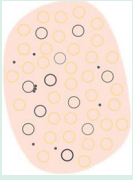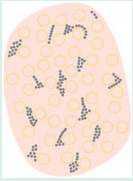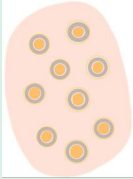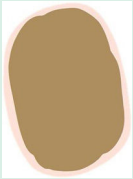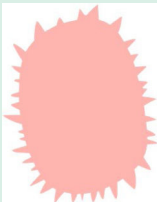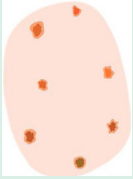
Criterion	Definition	Histopathological correlation	Schematic representation
Glomerular vessels	Larger caliber vessels than the dotted ones, convoluted or "coiled" resembling the renal glomerulus, often distributed in aggregates	Clustered and dilated capillaries in papillary dermis	
"Hairpin" vessels	Loop-shaped vessels in an oblique arrangement to skin surface, usually surrounded by a whitish halo in keratinizing tumors	Capillary loops in the papillary dermis in thick tumors	
Linear irregular	Linearly or slightly curved vessels of irregular shape, size, and/or distribution	Tumoral neoangiogenesis	
Polymorphous vascular pattern	Vessels of various morphologies, often including "hairpin," irregular linear, and dotted/glomerular vessels in invasive squamous cell carcinoma	Tumoral neoangiogenesis	
Pigment-associated criteria			
Brown follicular dots and circles	Small brown rings inside follicular openings	Presence of melanin in basal follicular cells at infundibular level	
Gray follicular dots and circles	Small gray rings inside follicular openings	Presence of melanin at isthmic level along with melanophages in the adjacent dermis	
Rhomboidal structures	Confluent gray dots in linear arrangement of gray-to-brown color between follicular openings	Pigmented keratinocytes in the Malpighian layer and/or melanophages in the superficial dermis	

Table 1 (*Continued*)
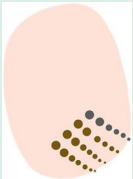
| Criterion	Definition	Histopathological correlation	Schematic representation
Inner gray halo	Subtle homogeneous gray or beige halo around follicular openings, forming an inner ring in relation to the pigmented pseudo-reticulum network	Area of preserved epidermis around follicular openings, with gray color as a resulta of the ''Tyndall effect'' of pigmented keratinocytes beneath this normal epidermis	
Brown structureless pigmentation	Homogeneous brown areas without other dermoscopic structures	Diffusely distributed melanin in basal keratinocytes	
Patchy or brown/gray dots/globules arranged in lines	Brown or gray dots arranged in a patchy or linear way, often found at the lesion periphery with radial orientation	Melanophages in dermal papillae near papillary vessels along with thin suprapapillary epidermis and/or increased number of pigmented keratinocytes	
Other criteria			
''Red starburst''	Presence of radial red lines or ''hairpin'' vessels around the central structureless white-yellow area of the lesion	May represent a sign of ''horizontal'' lesion growth	
Erosions	Small irregularly arranged structureless orange-to-red or brown areas	Loss of epidermis	
Ulceration	Large irregular or rounded structureless red or reddish-brown areas	Loss of epidermis and superficial dermis	

Source: Kittler et al.[9], Zalaudek et al.[10], Akay et al.[23], Lallas et al.[24], Ertop Doğan et al.[25], Nascimento et al.[26], Martín et al.[27], Zalaudek et al.[28], Zalaudek et al.[30], Cameron et al.[37], Rosendahl et al.[41], Yélamos et al.[52], and Kreusch[53].

mentation in 23.5% and 11.7% of cases, respectively[20], while Kelati et al. found an annular/granular pattern and rhomboidal structures in 19.4% and 82.8% of 232 pAKs.[22] However, certain dermoscopic criteria can aid in this difficult differential diagnosis. Lallas et al. studied a total of 70 LMs and 56 pAKs to find that the presence of white circles/prominent follicles (OR, 13.5; p = 0.006), scales (OR, 7.7; p = 0.001), and erythema (OR, 3.6; p = 0.009) correlated with the diagnosis of pAK. Conversely, rhomboidal structures, intense pigmentation, and non-prominent follicles were predictors of LM.[24]

A recent study including 53 pAKs confirmed that erythema (35.8%), scales (77.4%), and prominent follicles (52.8%) could be key in identifying these lesions while also seeing structures shared with LM such as brown dots (22.6%) and circles (43.4%), gray dots (45.4%) and circles (26.4%), and structureless pigmented areas (30.2%).[25] Finally, the presence of an ''inner gray halo'' at follicular level was reported by Nascimento et al. as a predictor of pAK vs LM, reporting this dermoscopic sign in 91.4% of pAKs vs 23.8% of LMs (p < 0.01), with excellent inter-observer agreement (κ = 0.846).[26]
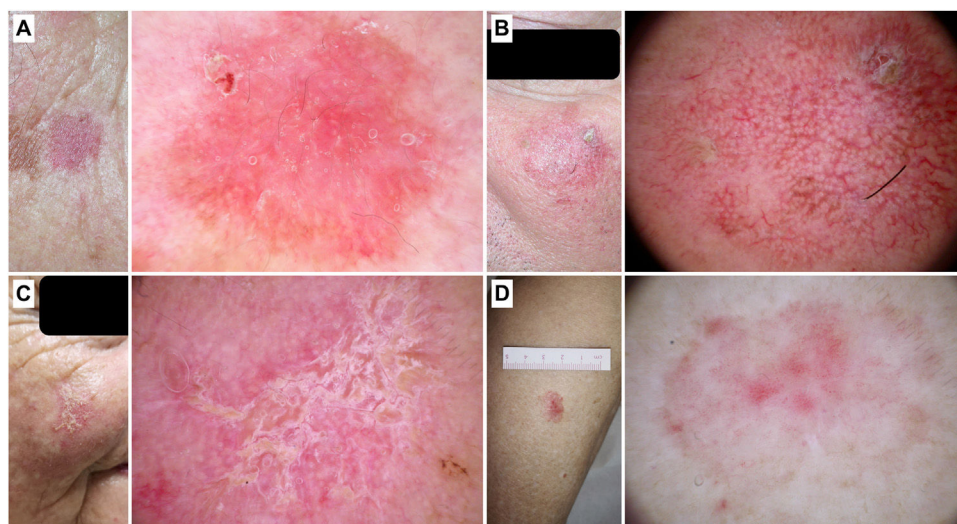
Figure 2 Non-pigmented actinic keratoses. A) Grade 1 actinic keratosis on the face of an 81-year-old woman. Dermoscopy shows a red pseudonetwork and discrete scaling. B) Grade 2 facial actinic keratosis in a 72-year-old man. ''Strawberry pattern,'' with prominent follicles/''rosettes,'' perifollicular wavy vessels, and keratin. C) Grade 3 actinic keratosis on the face of an 80-year-old woman. Dermoscopy shows keratin masses as white-yellowish structureless areas. D) Bowenoid actinic keratosis on the leg of an 80-year-old woman. Dermoscopy shows multiple dotted/glomerular vessels.

Other variants

Bowenoid AKs exhibit glomerular vessels of regular distributtion[27,28] (Fig. 2), while lichenoid AKs can exhibit a gray annular/granular pattern due to melanophagia.[5] Recently, the ''iceberg sign'' was described in AKs with a bluish surface coloration. The presence of this sign has been associated with the use of certain violet shampoos and the following deposition of amorphous basophilic material in the stratum corneum.[29]

Dermoscopy of in situ squamous cell carcinoma (Bowen's disease)

Non-pigmented Bowen's disease

Key aspects

Non-pigmented BD is characterized by the presence of clusters of glomerular or dotted vessels along with white/yellowish scales (Fig. 4).

In the initial study by Zalaudek et al. that analyzed 21 cases of BD found that the most common dermoscopic structures were glomerular vessels (90%) and white/yellowish scales (90%).[30] These data were corroborated by the same group, observing these criteria in 43.3% and 83.1% of all 71 cases of BD, with significant differences vs AK. The authors also described the presence of a ''red starburst'' pattern in 28.2% of BD cases. Based on the AK-BD-SCC progression model, they hypothesized that this criterion could represent an initial step in such progression.[10] In this regard, Pan et al. conducted a retrospective observational study with 50 cases of BD, 150 basal cell carcinomas (BCC), and 100 cases of psoriasis, concluding that the combination of clustered glomerular vessels and hyperkeratosis achieved a diagnostic likelihood of 98% for BD.[31] Of note that vessel morphology may be influenced by the magnification used.

Thus, with standard handheld dermatoscopes, we can see clusters of dotted vessels.[32] In a recent study, Papageorgiou et al. observed that dotted and glomerular vessels were the main predictors of BD vs BCC. However, they also noted that dotted or glomerular vessels can be detected in BCCs located on the lower limbs, likely due to venous stasis (25% and 19.3% in the study, respectively).[33] Aside from these fundamental findings, other described dermoscopic criteria include hemorrhages, focal hypopigmentation, linear irregular vessels, or ''hairpin'' vessels, among others.[34–36] Additionally, dermoscopy seems useful at post-therapeutic follow-up with imiquimod. In a small patient series, Mun et al. saw reported that persistent glomerular vessels after treatment would be suggestive of the presence of residual tumor.[34]

Pigmented Bowen's disease

Key aspects

Pigmented BD is characterized by the criteria present in the non-pigmented form and the presence of structureless brown pigmentation and brown or gray dots arranged in lines (Fig. 4).

In an initial study by Zalaudek's group that analyzed 10 pigmented BD cases and reported brown globules with a patchy distribution (90%), as well as structureless gray or brown areas (80%).[30] Afterwards, Cameron et al. published a retrospective study with 52 pigmented BD cases reporting the presence of brown or gray dots with a linear distribution pattern in 21.2% of cases. In 48.1% of cases, however, a structureless pigmentation pattern predominated, while 34.6% exhibited a combination of structureless pigmentation and dots. Most cases exhibited a monomorphic vascular pattern (82.9%), with predominance of glomerular vessels (44.2%), with a linear vessel distribution in 11.5%.[37] Other studies have corroborated these findings in varying percentages, mainly the presence of glomerular vessels (50% up to
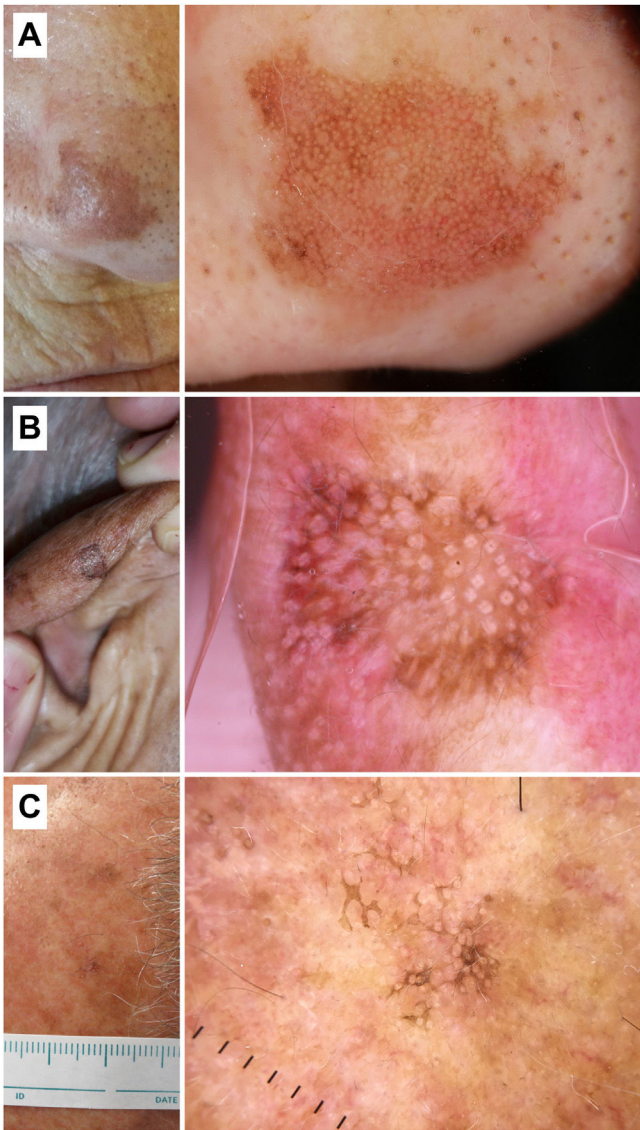
Figure 3 Pigmented actinic keratoses. A) Pigmented actinic keratosis on the nose of a 70-year-old woman. Dermoscopy shows an annular/granular pattern, erythema, and prominent follicles. B) Pigmented actinic keratosis on the helix of an 85-year-old man. Dermoscopy shows a structureless brown area, erythema, and multiple ''rosettes.'' C) Pigmented actinic keratosis on the temple of a 56-year-old man. Dermoscopy shows pigmented scales and prominent follicles/''rosettes.''

100%), pigmented dots/globules (30% up to 80%), or pigmented structureless areas (70% up to 78%).[34,36,38]

While these findings have proven reproducible in the genital area[38] (Fig. 4), a study that analyzed a total of 79 head and neck lesions concluded that dermoscopic patterns in this location differ from previously published data and are similar to pAKs based on a lower presence of glomerular vessels (7.6%) and dots with linear arrangement (13.9%), the observation of structures, such as pigmented circles (48.1%) and white circles (17.7%), rhomboidal structures (41.8%), and structureless pigmentation areas (86.1%), as well as the predominance of linear irregular (29.2%)[39] (Fig. 5). Other less consistently described dermoscopic criteria include projections, pigment network, hypopigmented areas, or ulceration[30,34,36,39].

Dermoscopy of invasive squamous cell carcinoma

Key aspects

In iSCC, we can identify dermoscopic criteria associated with keratinization (white clods and circles, white structureless areas), vascular criteria (linear irregular vessels, dotted/glomerular vessels, and ''hairpin'' vessels), and other criteria (ulceration, hemorrhages).

In well-to-moderately differentiated SCC, keratin-associated criteria are predominant, while in poorly differentiated SCC, a polymorphous vascular pattern predominates (Fig. 6).

In the differential diagnosis with AK, the presence of dotted/glomerular vessels, ''red starburst,'' ''hairpin'' vessels, white structureless areas, and perifollicular white circles should prompt a skin biopsy to rule out iSCC.

The study by Zalaudek et al. of 78 iSCCs and 24 KAs reported that ''hairpin'' vessels (38.5%), linear irregular vessels (17.9%), targetoid (41%), white structureless areas (42.3%), central keratin (39.4%), and ulceration (17.9%) were significantly associated with the diagnosis of iSCC (p < 0.001), with similar frequencies in the KA group except for linear irregular vessels, which were more common in the latter.[10] On the other hand, Jaimes et al. coined the term ''keratin pearls'' after studying a total of 15 well-differentiated KAs/SCCs and observed that all lesions exhibited rounded white/yellowish structures surrounded by a whitish halo[40] (Fig. 7). These structures, also known as white clods, were present in 25.6% and 16.7% of KA and SCC cases in the study by Rosendahl et al., respectively. This study, designed as a retrospective and prospective study with 43 KAs/60 iSCCs and 29 KAs/32 iSCCs/145 other lesions, respectively, concluded: 1) central keratin was more common in KA (51.2%) vs iSCC (30.0%) (p = 0.03); 2) the presence of keratin was more common in the KA/iSCC group vs other lesions (78.7% vs 30.3%; p < 0.001), with a sensitivity rate and positive predictive value (PPV) of 79% and 92% vs the BCC group, respectively; 3) white structureless areas (39.3% vs 18.6%; p = 0.02) and white circles (44.3% vs 13.1%; p < 0.001) were more common in the KA/iSCC group, with an 87% specificity rate for the latter vs other lesions; and 4) in the multivariate model, keratin, hemorrhages, and white circles were the only independent predictors of KA/iSCC diagnosis, with the latter reaching the highest ORs atv6.1 (95%CI, 2.4-13.3; p < 0.001).[41] In this context, of note that iSCC and KA can exhibit overlapping dermoscopic patterns, meaning that histopathological examinations will be necessary in most cases.[42]

Based on the degree of tumor differentiation, several dermoscopic patterns can be observed.[43,44] In summary, well-to-moderately differentiated iSCCs will more frequently exhibit ''hairpin'' vessels, white or yellowish structureless, white clods, and circles. Specifically, KA has been associated with the presence of a central keratin mass surrounded by ''hairpin'' or linear irregular with a radial distribution.[10,28] Conversely, poorly differentiated tumors will predominantly exhibit a polymorphous/atypical vascular pattern and ulceration.[5] Lallas et al. found that the
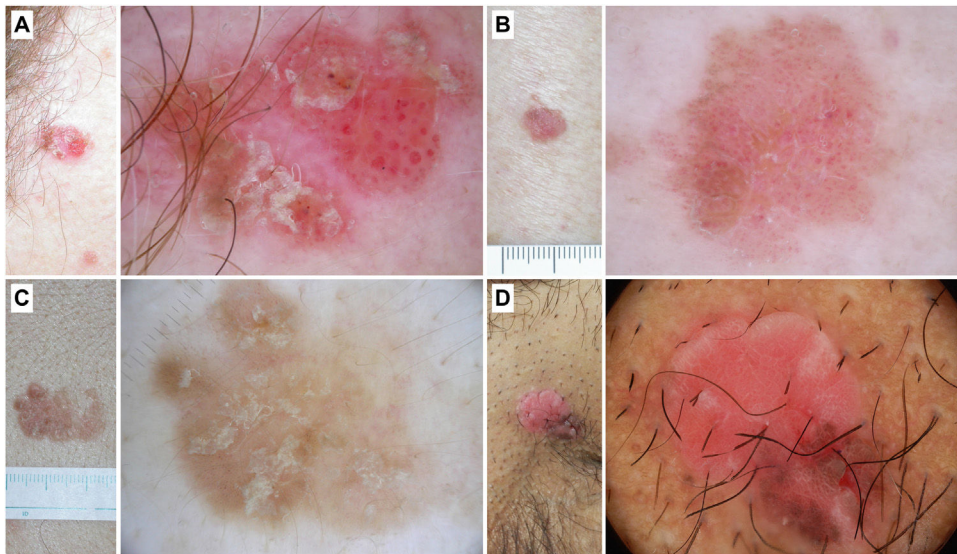
Figure 4 Bowen's disease. A) Non-pigmented Bowen's disease on the temple of a 79-year-old woman. Dermoscopic pattern consists of white-yellowish scales and clustered glomerular vessels. B) Non-pigmented Bowen's disease on the back of a 55-year-old man. Dermoscopy shows discrete scaling and glomerular and dotted vessels. C) Pigmented Bowen's disease on the back of a 31-year-old man. Dermoscopy shows structureless brown pigmentation, clustered glomerular vessels, and brown and gray dots arranged in lines. D) Genital Bowen's disease on the pubis of a 59-year-old man. Dermoscopy shows a structureless brown pigmentation area, brown dots arranged in lines, dotted vessels, and shiny white lines.
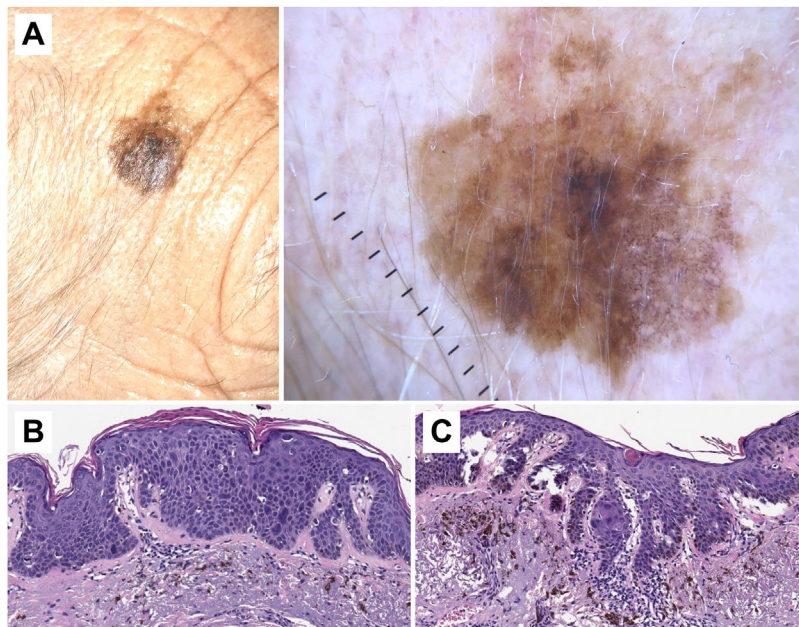
Figure 5 Histopathological correlation in pigmented actinic keratosis with areas of in situ squamous cell carcinoma. A) Pigmented atypical lesion on the temple of a 75-year-old woman. Dermoscopic image shows homogeneous brown pigmentation regions, brown circles, blue-gray granules, and gray dots arranged in lines in the lower part of the lesion. In this case, dermoscopic findings are not suitable to establish a reliable diagnosis, making histopahological examination essential. B) Histopathological image of an area of in situ squamous cell carcinoma (Bowen's disease) with melanophages on the superficial dermis (hematoxylin and eosin, ×10). C) Area of proliferative actinic keratosis with pigmented keratinocytes and melanophages on the superficial dermis (hematoxylin and eosin, ×10).
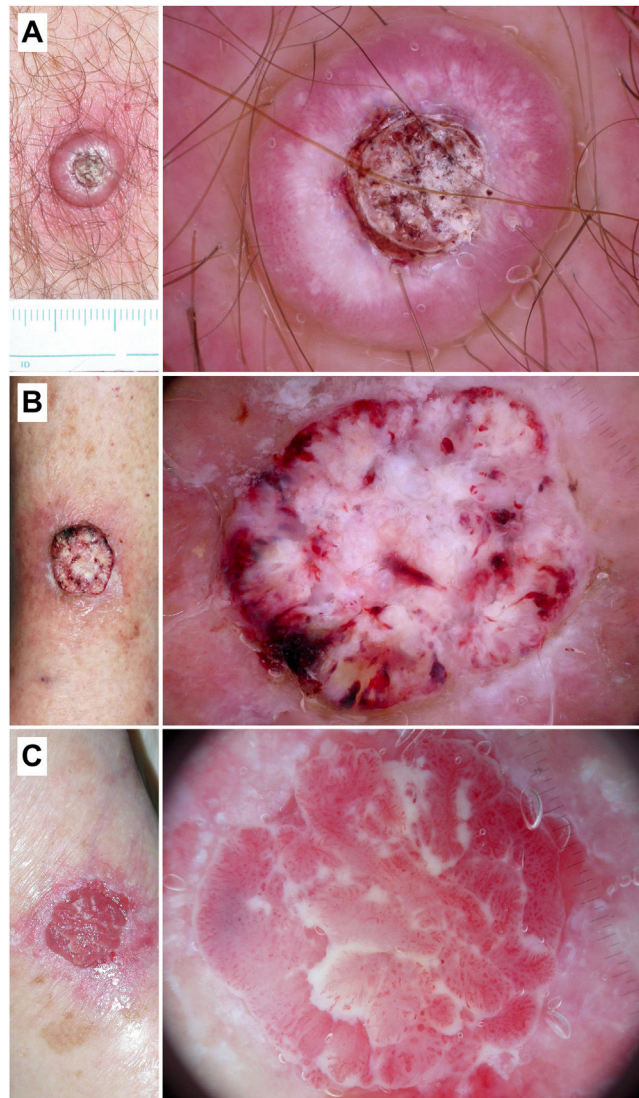
Figure 6 Keratoacanthoma and invasive squamous cell carcinoma. A) Keratoacanthoma on the chest of a 54-year-old man. Dermoscopy shows a central keratin mass, white clods and circles, white structureless areas, and ''hairpin'' vessels with a radial distribution. B) Well-differentiated squamous cell carcinoma on the leg of an 88-year-old woman. Dermoscopy shows white-yellowish structureless areas, white clods and circles, a polymorphous vascular pattern, and hemorrhages. C) Poorly differentiated squamous cell carcinoma on the arm of an 89-year-old woman. Dermoscopy shows a predominance of red color and an overtly atypical vascular pattern, with few keratin-associated criteria.

predominance of red color was associated with a 13-fold greater likelihood of being a poorly differentiated iSCC.[44] A similar pattern with predominant atypical vascularization has been reported in combined SCC/Merkel cell carcinoma tumors[45].

Regarding locations, we should mention that lip SCC shares most dermoscopic characteristics with cutaneous SCC.[46] Benati et al. published a series of cases of 22 lip SCC, in which the most relevant structures were scales (100%), perivascular white halos (86%), white structureless areas (91%), white circles (59%), and a polymorphous vascular pattern (68%).[47] In a recent multicentric retrospective study of 177 lip lesions (107 of them SCC), Lallas et al. saw that the presence of white clods and ulceration were predictors of SCC diagnosis vs controls (OR, 6.38 and 4.11, respectively).[48]

On the differential diagnosis with other lesions, early detection of iSCC and its differentiation from AK is essential, a common scenario in the follow-up of patients with actinic damage. The ''red starburst'' described by the study of Zalaudek et al. was observed in 29.5% of iSCC cases, with no significant differences being reported in frequency vs BD/in situ SCC. In any case, this dermoscopic pattern should be considered when planning to perform a skin biopsy in the context of a patient with actinic damage.[10] Papageorgiou et al. collected 50 incipient cases of iSCC and 45 AK with histopathological confirmation and found that the presence of dotted/glomerular vessels (OR, 3.83), ''hairpin'' vessels (OR, 12.12), and white structureless areas (OR, 3.58) were the main predictors for SCC diagnosis on the multivariate analysis. The univariate model also suggested that since
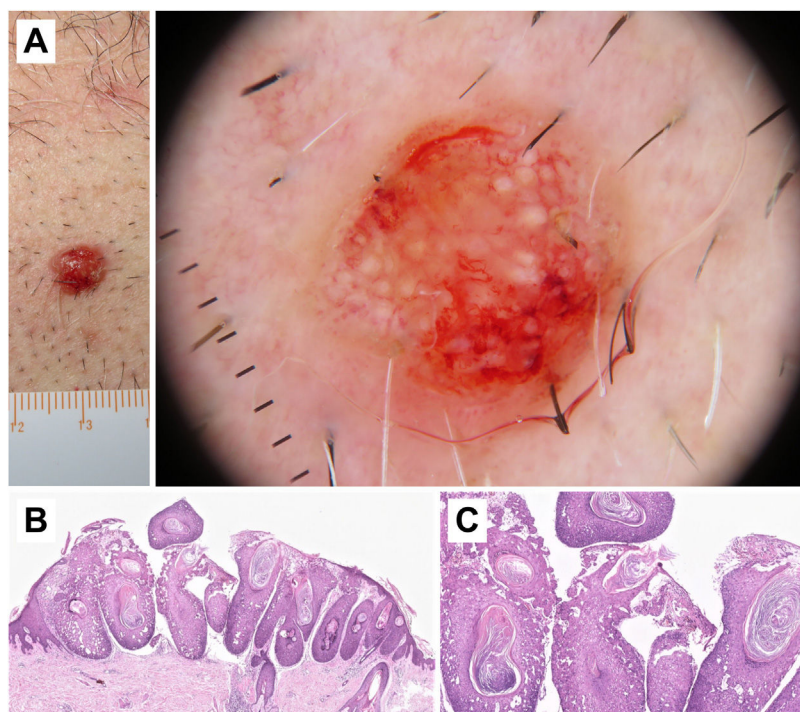
Figure 7 Histopathological correlation in microinvasive squamous cell carcinoma. A) Erythematous papule on the chest of a 46-year-old man. B) Dermoscopic image shows white clods and a polymorphous vascular pattern with linear irregular, ''hairpin,'' and glomerular vessels. C) Histopathological image shows epithelial proliferation with acanthosis and bulbous nests, revealing acantholysis and central keratinization phenomena (hematoxylin and eosin, ×2). D) Higher magnification image showing tumor cells and keratinization areas (''horn pearls''), corresponding to the white clods seen in dermoscopy (hematoxylin and eosin, ×5).

ulceration, perivascular white halos, and white circles could be predictors of SCC, therefore, they should also be taken into consideration.[49] iSCC can also significantly overlap with common benign lesions, such as irritated seborrheic keratosis (ISK), particularly in the case of well-differentiated SCC.[50] A study conducted by the same group analyzed 104 cases of SCC and 61 ISK and observed that the presence of dotted vessels (OR, 10.4), branched linear vessels (OR, 5.3), white structureless areas (OR, 6.78), white circles (OR, 23.45), or an irregular (OR, 2.55) or peripheral (OR, 2.8) distribution of vascular structures were predictors of SCC diagnosis vs ISK.[51]

Conclusions

Patients with actinic damage/field cancerization generally present with several skin lesions of varying biological behavior. In this context, an accurate differential diagnosis that allows us to select malignant lesions amenable to surgery and reliably identify ''premalignant'' lesions amenable to other treatments is desirable. Based on the available evidence, dermoscopy can be key in this endeavor, and therefore, appropriate training beyond melanocytic neoplasms, integrating the spectrum of keratinocytic carcinomas, is essential.

Conflicts of interest

None declared.

Acknowledgments

References

1. Que SKT, Zwald FO, Schmults CD. Cutaneous squamous cell carcinoma: Incidence, risk factors, diagnosis, and staging. J Am Acad Dermatol. 2018;78:237–47, http://dx.doi.org/10.1016/J.JAAD.2017.08.059.

2. Stratigos AJ, Garbe C, Dessinioti C, Lebbe C, Bataille V, Bastholt L, et al. European interdisciplinary guideline on invasive squamous cell carcinoma of the skin: Part 1. Epidemiology, diagnostics and prevention. Eur J Cancer. 2020;128:60–82, http://dx.doi.org/10.1016/J.EJCA.2020.01.007.

3. Tejera-Vaquerizo A, Descalzo-Gallego MA, Otero-Rivas MM, Posada-García C, Rodríguez-Pazos L, Pastushenko I, et al. Skin Cancer Incidence and Mortality in Spain: A Systematic Review and Meta-Analysis. Actas Dermosifiliogr. 2016;107:318–28, http://dx.doi.org/10.1016/J.AD.2015.12.008.

4. Lin MJ, Xie C, Pan Y, Jalilian C, Kelly JW. Dermoscopy improves diagnostic accuracy for clinically amelanotic nodules. Australas J Dermatol. 2019;60:45–9, http://dx.doi.org/10.1111/AJD.12902.

5. Zalaudek I, Argenziano G. Dermoscopy of actinic keratosis, intraepidermal carcinoma and squamous cell carcinoma. Curr Probl Dermatol. 2015;46:70–6, http://dx.doi.org/10.1159/000366539.

6. Zaballos Diego P. Dermoscopy Can Indicate the Grade of Actinic Keratoses. Actas Dermosifiliogr. 2017;108:798, http://dx.doi.org/10.1016/J.AD.2017.07.005.

7. Valdés-Morales KL, Peralta-Pedrero ML, Jurado-Santa Cruz F, Morales-Sánchez MA. Diagnostic Accuracy of Dermoscopy of Actinic Keratosis: A Systematic Review. Dermatol Pract Concept. 2020;10, http://dx.doi.org/10.5826/DPC.1004A121.

8. Zalaudek I, Giacomel J, Argenziano G, Hofmann-Wellenhof R, Micantonio T, et al. Dermoscopy of facial nonpigmented actinic keratosis. Br J Dermatol. 2006;155:951–6, http://dx.doi.org/10.1111/J.1365-2133.2006.07426.X.

9. Kittler H, Marghoob AA, Argenziano G, Carrera C, Curiel-Lewandrowski C, Hofmann-Wellenhof R, et al. Standardization of terminology in dermoscopy/dermatoscopy: Results of the third consensus conference of the International Society of Dermoscopy. J Am Acad Dermatol. 2016;74:1093, http://dx.doi.org/10.1016/J.JAAD.2015.12.038.

10. Zalaudek I, Giacomel J, Schmid K, Bondino S, Rosendahl C, Cavicchini S, et al. Dermatoscopy of facial actinic keratosis, intraepidermal carcinoma, and invasive squamous cell carcinoma: A progression model. J Am Acad Dermatol. 2012;66:589–97, http://dx.doi.org/10.1016/J.JAAD.2011.02.011.

11. Cuellar F, Vilalta A, Puig S, Palou J, Salerni G, Malvehy J. New dermoscopic pattern in actinic keratosis and related conditions. Arch Dermatol. 2009;145:732, http://dx.doi.org/10.1001/ARCHDERMATOL.2009.86.

12. Liebman TN, Scope A, Rabinovitz H, Braun RP, Marghoob AA. Rosettes may be observed in a range of conditions. Arch Dermatol. 2011;147:1468, http://dx.doi.org/10.1001/ARCHDERMATOL.2011.312.

13. Lozano-Masdemont B, Polimón-Olabarrieta I, Marinero-Escobedo S, Gutiérrez-Pecharromán A, Rodríguez-Lomba E. Rosettes in actinic keratosis and squamous cell carcinoma: Distribution, association to other dermoscopic signs and description of the rosette pattern. J Eur Acad Dermatol Venereol. 2018;32:48–52, http://dx.doi.org/10.1111/JDV.14474.

14. Çayirli M, Köse O, Demiriz M. Dermatoscopy of facial nonpigmented actinic keratosis regarding to grades of the lesions. J Eur Acad Dermatol Venereol. 2013;27:1185–7, http://dx.doi.org/10.1111/JDV.12020.

15. Reinehr CPH, Garbin GC, Bakos RM. Dermatoscopic Patterns of Nonfacial Actinic Keratosis: Characterization of Pigmented and Nonpigmented Lesions. Dermatol Surg. 2017;43:1385–91, http://dx.doi.org/10.1097/DSS.0000000000001210.

16. Huerta-Brogeras M, Olmos O, Borbujo J, Hernández-Núñez A, Castaño E, Romero-Maté A, et al. Validation of dermoscopy as a real-time noninvasive diagnostic imaging technique for actinic keratosis. Arch Dermatol. 2012;148:1159–64, http://dx.doi.org/10.1001/ARCHDERMATOL.2012.1060.

17. Pampena R, Condorelli A, Cornacchia L, Guiducci L, Raucci M, Benati E, et al. Treatment monitoring of 5-fluorouracil 0.5%/salicylic acid 10% lesion-directed therapy for actinic keratosis using dermoscopy and in-vivo reflectance confocal microscopy. Dermatol Ther. 2020;33, http://dx.doi.org/10.1111/DTH.13744.

18. Lee JH, Won CY, Kim GM, Kim SY. Dermoscopic features of actinic keratosis and follow up with dermoscopy: A pilot study. J Dermatol. 2014;41:487–93, http://dx.doi.org/10.1111/1346-8138.12282.

19. Zalaudek I, Ferrara G, Leinweber B, Mercogliano A, D'Ambrosio A, Argenziano G. Pitfalls in the clinical and dermoscopic diagnosis of pigmented actinic keratosis. J Am Acad Dermatol. 2005;53:1071–4, http://dx.doi.org/10.1016/J.JAAD.2005.08.052.

20. Moscarella E, Rabinovitz H, Zalaudek I, Piana S, Stanganelli I, Oliviero MC, et al. Dermoscopy and reflectance confocal microscopy of pigmented actinic keratoses: A morphological study. J Eur Acad Dermatol Venereol. 2015;29:307–14, http://dx.doi.org/10.1111/JDV.12532.

21. Stolz W, Schiffner R, Burgdorf WHC. Dermatoscopy for facial pigmented skin lesions. Clin Dermatol. 2002;20:276–8, http://dx.doi.org/10.1016/S0738-081X(02)00221-3.

22. Kelati A, Baybay H, Moscarella E, Argenziano G, Gallouj S, Mernissi FZ. Dermoscopy of Pigmented Actinic Keratosis of the Face: A Study of 232 Cases [Article in English, Spanish]. Actas Dermosifiliogr. 2017;108:844–51, http://dx.doi.org/10.1016/J.AD.2017.05.002.

23. Akay BN, Kocyigit P, Heper AO, Erdem C. Dermatoscopy of flat pigmented facial lesions: Diagnostic challenge between pigmented actinic keratosis and lentigo maligna. Br J Dermatol. 2010;163:1212–7, http://dx.doi.org/10.1111/J.1365-2133.2010.10025.X.

24. Lallas A, Tschandl P, Kyrgidis A, Stolz W, Rabinovitz H, Cameron A, et al. Dermoscopic clues to differentiate facial lentigo maligna from pigmented actinic keratosis. Br J Dermatol. 2016;174:1079–85, http://dx.doi.org/10.1111/BJD.14355.

25. Ertop Doğan P, Akay BN, Okçu Heper A, Rosendahl C, Erdem C. Dermatoscopic findings and dermatopathological correlates in clinical variants of actinic keratosis, Bowen's disease, keratoacanthoma, and squamous cell carcinoma. Dermatol Ther. 2021;34, http://dx.doi.org/10.1111/DTH.14877.

26. Nascimento MM, Shitara D, Enokihara MMSS, Yamada S, Pellacani G, Rezze GG. Inner gray halo, a novel dermoscopic feature for the diagnosis of pigmented actinic keratosis: Clues for the differential diagnosis with lentigo maligna. J Am Acad Dermatol. 2014;71:708–15, http://dx.doi.org/10.1016/J.JAAD.2014.05.025.

27. Martín JM, Bella-Navarro R, Jordá E. Vascular patterns in dermoscopy [Article in Spanish]. Actas Dermosifiliogr. 2012;103:357–75, http://dx.doi.org/10.1016/J.AD.2011.11.005.

28. Zalaudek I, Kreusch J, Giacomel J, Ferrara G, Catrical C, Argenziano G. How to diagnose nonpigmented skin tumors: A review of vascular structures seen with dermoscopy: Part II. Nonmelanocytic skin tumors. J Am Acad Dermatol. 2010;63:377–86, http://dx.doi.org/10.1016/J.JAAD.2009.11.697.

29. Mir-Bonafé JF, Mir-Bonafé M, Mozos A, López-Sánchez C, Piquero-Casals J, de Dios-Velázquez Á, et al. Iceberg sign in actinic keratosis neglecta caused by toning shampoo for blonde and white hair. Int J Dermatol. 2020;59:e64–5, http://dx.doi.org/10.1111/IJD.14717.

30. Zalaudek I, Argenziano G, Leinweber B, Citarella L, Hofmann-Wellenhof R, Malvehy J, et al. Dermoscopy of Bowen's disease. Br J Dermatol. 2004;150:1112–6, http://dx.doi.org/10.1111/J.1365-2133.2004.05924.X.

31. Pan Y, Chamberlain AJ, Bailey M, Chong AH, Haskett M, Kelly JW. Dermatoscopy aids in the diagnosis of the solitary red scaly patch or plaque-features distinguishing superficial basal cell carcinoma, intraepidermal carcinoma, and psoriasis. J Am Acad Dermatol. 2008;59:268–74, http://dx.doi.org/10.1016/J.JAAD.2008.05.013.

32. Bugatti L, Filosa G, De Angelis R. The specific dermoscopical criteria of Bowen's disease. J Eur Acad Dermatol Venereol. 2007;21:700–1, http://dx.doi.org/10.1111/J.1468-3083.2006.01995.X.

33. Papageorgiou C, Apalla Z, Variaah G, Matiaki FC, Sotiriou E, Vakirlis E, et al. Accuracy of dermoscopic criteria for the differentiation between superficial basal cell carcinoma and Bowen's disease. J Eur Acad Dermatol Venereol. 2018;32:1914–9, http://dx.doi.org/10.1111/JDV.14995.

34. Mun JH, Kim SH, Jung DS, Ko HC, Kwon KS, Kim MB. Dermoscopic features of Bowen's disease in Asians. J Eur Acad Dermatol Venereol. 2010;24:805–10, http://dx.doi.org/10.1111/J.1468-3083.2009.03529.X.

35. Yang Y, Lin J, Fang S, Han S, Song Z. What's new in dermoscopy of Bowen's disease: Two new dermoscopic signs and its differential diagnosis. Int J Dermatol. 2017;56:1022–5, http://dx.doi.org/10.1111/IJD.13734.

36. Payapvipapong K, Tanaka M. Dermoscopic classification of Bowen's disease. Australas J Dermatol. 2015;56:32–5, http://dx.doi.org/10.1111/AJD.12200.

37. Cameron A, Rosendahl C, Tschandl P, Riedl E, Kittler H. Dermatoscopy of pigmented Bowen's disease. J Am Acad Dermatol. 2010;62:597–604, http://dx.doi.org/10.1016/J.JAAD.2009.06.008.

38. Giuffrida R, Conforti C, Resende FSS, Hamilko de Barros M, Uranitsch M, Favero F, et al. Clinical and dermoscopic features of genital pigmented Bowen disease. Clin Exp Dermatol. 2018;43:813–6, http://dx.doi.org/10.1111/ced.13633.

39. Inskip M, Cameron A, Akay BN, Gorji M, Clark SP, Rosendahl N, et al. Dermatoscopic features of pigmented intraepidermal carcinoma on the head and neck. J Dtsch Dermatol Ges. 2020;18:969–76, http://dx.doi.org/10.1111/DDG.14220.

40. Jaimes N, Zalaudek I, Braun RP, Tan BH, Busam KJ, Marghoob AA. Pearls of keratinizing tumors. Arch Dermatol. 2012;148:976, http://dx.doi.org/10.1001/ARCHDERMATOL.2011.3475.

41. Rosendahl C, Cameron A, Argenziano G, Zalaudek I, Tschandl P, Kittler H. Dermoscopy of squamous cell carcinoma and keratoacanthoma. Arch Dermatol. 2012;148:1386–92, http://dx.doi.org/10.1001/ARCHDERMATOL.2012.2974.

42. Sgouros D, Theofili M, Damaskou V, Theotokoglou S, Theodoropoulos K, Stratigos A, et al. Dermoscopy as a Tool in Differentiating Cutaneous Squamous Cell Carcinoma From Its Variants. Dermatol Pract Concept. 2021;11, http://dx.doi.org/10.5826/DPC.1102A50.

43. Manfredini M, Longo C, Ferrari B, Piana S, Benati E, Casari A, et al. Dermoscopic and reflectance confocal microscopy features of cutaneous squamous cell carcinoma. J Eur Acad Dermatol Venereol. 2017;31:1828–33, http://dx.doi.org/10.1111/JDV.14463.

44. Lallas A, Pyne J, Kyrgidis A, Andreani S, Argenziano G, Cavaller A, et al. The clinical and dermoscopic features of invasive cutaneous squamous cell carcinoma depend on the histopathological grade of differentiation. Br J Dermatol. 2015;172:1308–15, http://dx.doi.org/10.1111/BJD.13510.

45. Suárez AL, Louis P, Kitts J, Busam K, Myskowski PL, Wong RJ, et al. Clinical and dermoscopic features of combined cutaneous squamous cell carcinoma (SCC)/neuroendocrine [Merkel cell] carcinoma (MCC). J Am Acad Dermatol. 2015;73:968–75, http://dx.doi.org/10.1016/J.JAAD.2015.08.041.

46. Güleç AT. Diagnosing squamous cell carcinoma of the lip using dermoscopy. J Am Acad Dermatol. 2017;76:S82–3, http://dx.doi.org/10.1016/J.JAAD.2016.10.026.

47. Benati E, Persechino F, Piana S, Argenziano G, Lallas A, Moscarella E, et al. Dermoscopic features of squamous cell carcinoma on the lips. Br J Dermatol. 2017;177:e41–3, http://dx.doi.org/10.1111/BJD.15274.

48. Lallas A, Martínez G, Arceu M, Kyrgidis A, Liopyris K, Brancaccio G, et al. Clinical and dermatoscopic predictors of squamous cell carcinoma of the lips: A case-control, multicentric study. J Eur Acad Dermatol Venereol. 2022;36:222–7, http://dx.doi.org/10.1111/JDV.17790.

49. Papageorgiou C, Lallas A, Manoli SM, Longo C, Lai M, Liopyris K, et al. Evaluation of dermatoscopic criteria for early detection of squamous cell carcinoma arising on an actinic keratosis. J Am Acad Dermatol. 2022;86:791–6, http://dx.doi.org/10.1016/J.JAAD.2021.03.111.

50. Papageorgiou V, Apalla Z, Sotiriou E, Papageorgiou C, Lazaridou E, Vakirlis S, et al. The limitations of dermoscopy: False-positive and false-negative tumours. J Eur Acad Dermatol Venereol. 2018;32:879–88, http://dx.doi.org/10.1111/JDV.14782.

51. Papageorgiou C, Spyridis I, Manoli SM, Busila I, Nasturica IE, Lallas K, et al. Accuracy of dermoscopic criteria for the differential diagnosis between irritated seborrheic keratosis and squamous cell carcinoma. J Am Acad Dermatol. 2021;85:1143–50, http://dx.doi.org/10.1016/J.JAAD.2020.02.019.

52. Yélamos O, Braun RP, Liopyris K, Wolner ZJ, Kerl K, Gerami P, et al. Dermoscopy and dermatopathology correlates of cutaneous neoplasms. J Am Acad Dermatol. 2019;80:341–63, http://dx.doi.org/10.1016/j.jaad.2018.07.073.

53. Kreusch JF. Vascular patterns in skin tumors. Clin Dermatol. 2002;20:248–54, http://dx.doi.org/10.1016/s0738-081x(02)00227-4.